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(71) Applicant (for all designated States except US): MINISTERO UNIVERSITA' RICERCA SCIENTIFICA E TECNOLOG-ICA [IT/IT]; Piazzale Kennedy, 20, I-00144 Rome (IT).

(72) Inventors; and

- (75) Inventors/Applicants (for US only): MELE, Antonio [IT/IT]; Via E. Magnani, 16, I-51016 Montecatini Terme (IT). DE SANTIS, Rita [IT/IT]; Via Catilina, 14, I-00040 Pomezia (IT). PARENTE, Dino [IT/IT]; Via Plinio, 18, I-00040 Pomezia (IT). COLNAGHI, Maria, Ines [IT/IT]; Strada al Lago, 12, I-20090 Milan San Felice (IT).
- (74) Agent: GERVASI, Gemma; Notarbartolo & Gervasi, Corso di Porta Vittoria, 9, I-20122 Milan (IT).

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(54) Title: RECOMBINANT RIBOSOMAL INHIBITOR PROTEIN (RIP) AND USE AS IMMUNOCONJUGATE

(57) Abstract

The following description refers to a new RIP protein (SEQ ID No: 3) the cDNA sequence expressing same (SEQ ID No: 2), its preparation and use in the preparation of chemical and recombinant conjugates having anticancer properties.

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RECOMBINANT RIBOSOMAL INHIBITOR PROTEIN (RIP) AND USE AS IMMUNOCONJUGATE

Field of the invention

The present invention relates to a new protein which - after being transferred into the cell by a suitable vector - is capable of inhibiting the activity of ribosomes.

Therefore, it can be used as an anticancer and/or antiviral agent.

State of the art

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As known, proteins extracted from filamentous fungi of the genus Aspergillus, such as α -sarcin, restrictocin and mitogillin, can inhibit protein synthesis by inactivating eukaryotic ribosomes. The nucleotide sequence of genomic DNA expressing said proteins is also known.

The antiviral activity of RIPs is related to the higher membrane permeability to the RIPs of virus-infected cells, with consequent injury to their ribosomes and consequent death of the infected cell. It follows that viral replication is interrupted. It has recently been disclosed that several RIPs inhibit HIV replication and that a RIP preparation, thrichosantin (a protein extracted from the roots of *Trichosantes kiriloowii*) was used in phase I/II clinical studies [Byers, VS. et al., A phase I/II study of thrichosantin treatment of HIV diseases, AIDS, 4, 1189-1196 (1990)].

With a view to obtain selectively cytotoxic molecules, several of the known RIPs were bound to proteic and non-proteic vectors capable of trasferring them on specific target cell populations. Compounds with a specific cytotoxic action are most frequently prepared with monoclonal antibodies as protein vectors (immunotoxins). However, hormones, growth factors, lectins have also been used as vectors for the treatment of cancer.

The protein that has been most widely used so far for the construction of immunotoxins is ricin chain A; however, several RIPs of type 1 (gelonin, PAP, saporin, momordin, bryodin, barley RIP) have been recently tested in the treatment, e.g., of tumours, autoimmune diseases, transplant rejections, parasitoses, etc.

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Since tumour cells are often toxin-resistant and all toxins used so far in therapeutic treatments induce an immune response in treated patients, the identification and purification to homogeneity of new RIPs are of great importance in all therapeutic applications and in particular in the generation of new immunotoxins.

Summary of the invention

The present invention relates to a new RIP protein SEQ ID No: 3, able to inhibit protein synthesis by inactivating ribosomes. Said RIP protein is herein referred to as clavin.

The present invention also concerns the nucleotide sequence SEQ ID No: 2, responsible of the expression of the new RIP protein clavin.

The present invention further includes the conjugates of the aforesaid protein with monoclonal antibodies, hormones, liposomes, growth factors, cytokines, transferrin and peptides, consisting of fragments of said proteins, obtainable by chemical conjugation or by genic recombination techniques whenever applicable. It is another object of the present invention the Mgr6-clavin conjugate having amino acid composition SEQ ID No: 5, as well as the nucleotide sequence

Detailed description of the invention

expressing it, corresponding to SEQ ID No: 4.

- SEQ ID No: 1 reports the complete cDNA sequence (i.e., including non-coding 3' and 5' sequences) of clavin, in which:
 - 1-279 = 5' UTR (5' fragment, untranslated);
 - 280-360 = sequence encoding for hypothetical secretion sequence;
 - 361-813 = sequence encoding for mature protein (clavin);
- 25 814-1011 = 3' UTR (3' fragment, untranslated) + polyA.
 - SEQ ID No: 2 refers to the cDNA sequence encoding for mature clavin.
 - SEQ ID No: 3 corresponds to clavin protein sequence.
 - SEQ ID No: 4 describes the nucleotide sequence encoding for the Mgr6-clavin immunotoxin produced in pRSET, in which:
- 30 1-108 = pRSET sequence containing the 6 histidines and the cleavage site for enterokinase;

109-861 = sequence encoding for ScFv (single Fv) of Mgr6

(109-462 = variable sequence of heavy chain, Bankit I.D. (Gene Bank) 54241, access No. U 61494)

(463-507 = linker sequence)

5 (508-861 = variable sequence of light chain, Bankit I.D. (Gene Bank) 54263, access No. U 61495)

(862-1311 = sequence encoding for clavin).

SEQ ID IN No: 5 corresponds to the protein sequence of Mgr6-claim immunotoxin produced in pRSET

The purified protein is >95% pure as shown by SDS/PAGE analysis, N-terminal sequencing and reversed-phase HPLC. The protein molecular weight is approx. 17kDa.

The following examples are conveyed for a better understanding of the protein purification process according to the invention.

15 cDNA isolation and sequencing

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Total RNA was extracted from Aspergillus clavatus IFO 8605 (Institute of Fermentation, Osaka). mRNA was purified using the kits for total RNA and, respectively, mRNA purification (Clontech). The two following primers were synthesized:

3' α-primer: 5'-ACGTAAGCTTCTAATGAGAGCAGAGCTT-3' (SEQ ID No: 6) 5' α-primer: 5'-ACGTCTGCAGTGACCTGGACCTGCTTGAACG-3' (SEQ ID No: 7) Primers were drawn on the basis of the known α-sarcin sequence by assuming a high amino acid sequence homology with the toxin of *Aspergillus clavatus*.

The synthesis of cDNA with 5 μ g mRNA and 3' α -primer was carried out using an appropriate synthesis kit (BRL). Part of the product obtained was added to a reaction mixture for PCR containing Taq polymerase (USB) and the two aforesaid primers; cDNA amplification for a total of 30 cycles was performed using a thermal cyclizer for DNA (Perkin-Elmer).

Each cycle consisted of 1-min denaturation at 94°C, 1-min annealing at 42°C and 2-min extension at 72°C; in the final cycle extension at 72°C lasted 8 min.

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Amplified cDNA corresponded to the cDNA of clavin, but contained the sequences imposed by $3'\alpha$ - and $5'\alpha$ -primers. Isolation of complete native cDNA of clavin was carried out by the RACE method.

On 3' end, RACE was performed according to Frohman, using the following primer:

5'-GACTCGAGTCGACATCGA(T)₁₇-3' (SEQ ID No: 8), and the adjustment primer: 5'-GACTCGAGTCGACATCG-3' (SEQ ID No: 9).

Primer 5'-ACGTGGATCCTCTACAACCAGAAC-3' (SEQ ID No: 10), which refers to the codons for amino acids 23-29 of mature protein and bearing a restriction site BamHI, was used as a gene-specific primer.

On 5' end, RACE was performed using the 5'-AmpliFINDER RACE Kit (Clontech), and primers

5'-TGAACCAGTGAGGATAG-3' (SEQ ID No: 11)

5'-ACGTCTGCAGGCGCTTGTTCTCATA-3' (SEQ ID No: 12)

referring to the codons for amino acids 47-53 and 18-23 of mature protein were used as gene-specific primers; the latter primer also contains a restriction site Pstl.

The various PCR products were purified, digested and subcloned in pUC19. Sequeces were analysed using PC GENE software (Intelligenetics).

The complete cDNA sequence obtained is shown in Fig. 1. Said sequence contains an ORF encoding for a 177 amino acid polypeptide chain. The first 27 amino acids represent a signal peptide involved in secretion, while mature protein consists of the 150 amino acids shown in the figure.

Recombinant clavin heterologous expression

- Vector pEZZ18 (Pharmacia) was used for recombinant clavin heterologous expression. Said vector directs the expression of fused proteins with a linking synthetic domain IgG (ZZ) based on staphylococcus protein A (Nilsson *et al.*, 1987). Clavin cDNA obtained by PCR with primers based on α-sarcin, as previously described, was re-amplified with 3' primer:
- 30 5'-GATCCTGCAGCGACCTGGACTTGCATGAACGAGCAGAAGAACCCAAAG-ACC-3' (SEQ ID No: 13)

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and with 3' primer: 5'-ACGTAAGCTTCTAATGAGAGCAGAGCTT-3' (SEQ ID No: 14)

to obtain the mature clavin native sequence, and cloned at restriction sites Pstl-HindIII of vector pEZZ18. To obtain pMRS116, fragment EcoRI-Pstl was replaced by linker B, which contains a sequence encoding for the cleavage site of factor Xa Ile-Glu-Gly-Arg, in addition to a residue Thr, inserted to preserve restriction site Pstl. Linker B was obtained by annealing of the two oligonucleotides:

α-28: 5'-AATTCGATCGAAGGTCGTACTGCA-3' (SEQ ID No: 15)

α-29: 5'-GTACGACCTTCGATCG-3' (SEQ ID No: 16).

For clavin production, construction pMRS116 was propagated in *Escherichia coli* HB 101 [supE44, hsdS20(r_Bm_B)recA13, ara-14, proA2, lacY1, galK2, rpsL20, xyl-5, mtl-1] and cultured according to the producer's directions (Pharmacia).

Recombinant clavin purification

Culture supernatants were brought to pH 7.6 and added with 1 mM phenylmethylsulfonyl fluoride. Supernatants were injected into an IgG Sepharose Fast Flow column (Pharmacia) equalized with buffer (50 mM Tria/HCl, 150 mM NaCl and 0.05% Tween-20, pH 7.6). The column was first washed with said buffer and then with 5 mM ammonium acetate, pH 5.1; the fusion protein was eluted with 0.5 M ammonium acetate, pH 3.4, freeze-dried, dissolved to 2-8 mg/ml in 20 mM Tris/HCl, 100 mM NaCl, 1 mM CaCl₂₁ pH 8.0 and digested with Xa factor (Boehringer) at 23°C for 18-24 h in an enzyme/substrate ratio equal to 1:100. Clavin was purified using a two-phase chromatographic process. In the first phase, the digestion mixture was injected into an S Sepharose Fast Flow column (Pharmacia) equalized with 20 mM sodium phosphate, pH 5.8, and eluted in NaCl gradient; the fusion protein left undigested and clavin were eluted in a single peak. The second phase was still performed on IgG Sepharose Fast Flow column: clavin was collected with the eluent, while the non-digested protein was eluted from the colum with 0.5 M ammonium acetate, pH 3.4.

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Immunotoxin synthesis and purification (chemical conjugate)

Clavin was derivatized with 40-molar excess ethyl S-acetyl-3-propionthioimidate at 4°C for 1 h to obtain an average molar ratio of ethyl S-acetyl-3-propionthioimidate groups to toxin molecule equal to 1.2.

Monoclonal antobody Mgr6, directed against the extracellular domain of ErbB2 and produced from hybridoma Mgr6-C4 MCB c # 762, deposited in the Interlab Cell Line Collection bank (CBA), an international deposit authority, was purified from ascitic liquid as described in Centis *et al.*, 1992. It was then derivatized with 10-molar excess 2-iminothiolane in ethanol at room temperature for 30 min, added with 5,5'-dithio-bis(2-nitrobenzoic acid) to block the free -SH groups and obtain a groups/toxin molecule molar ratio equal to 1:6. The mixture was applied to a BioGel P-6DG column to remove all reagents.

Derivatized products were mixed using a 5-molar excess clavin, concentrated to a final volume of 1 ml, and added with 100 ul of 0.5 M hydroxylamine, 12.5 mM EDTA, pH 7.2. The solution was stirred at 22°C for 14 h and at 4°C for additional 18 h. The reaction was interrupted by addition of 20 ul of 200 mM N-ethylmaleimide.

The immunoconjugate was purified to homogeneity by ion exchange chromatography.

20 Recombinant immunotoxin Mgr6-clavin

Genic construction

The gene encoding for variable regions of monoclonal antobody Mgr6 was obtained by the "Recombinant Phase Antibody System" kit (Pharmacia) from mRNA of the antibody-producing hybridoma. Said procedure allows the obtainment of DNA encoding for ScFv (Single chain Fv), in which the sequences for the variable regions of heavy and light chains are joined by a linker sequence. The DNA for ScFv was then linked to clavin DNA to obtain the gene for immunotoxin, cloned in commercial vector pRSET (Introvigen). Figs. 4 and 5 show the nucleotide and, respectively, the amino acid sequence of the immunotoxin inserted in pRSET.

The resulting plasmid has the following characteristics:

- a) the fusion protein gene is under the control of the T7 polymerase control;
- b) the resulting protein has at its N-terminal site an extension containing 6 histidines, usable for the purification with IMAC (immobilized metal affinity chromatography) and a cleavage site for enterokinase K.

Recombinant immunotoxin expression in E. coli

Competent cells B834(DE3)pLys were transformed, plated on 10 LB agar plates containing ampicillin (100 µg/ml), and incubated at 37°C overnight.

Colonies were recovered in 500 ml LB culture medium containing ampicillin (100 μ g/ml), glucose (0.5%) and MgSO₄ (1,62 mM), cultured at 37°C under stirring up to OD₆₀₀ = 2.3/2.5, and, after addition of 1 l culture medium (LB, ampicillin, glucose, MgSO₄), amplified at 37°C under stirring up to OD₆₀₀ = 1.2.

Cells were centrifuged and resuspended in 1.5 I culture medium LB supplemented with ampicillin and induced by addition of IPTG (final 1 mM) under stirring at 37°C for 1.5 h. The cell pellet was recovered by centrifugation.

15 Recombinant immunotoxin purification

The pellet from 1.5 I culture medium was resuspended in 150 ml of 50 mM Tris-HCI, pH 8, and frozen. 30 ml aliquots were thawed out, sonicated (3 \times 20 sec), and centrifuged at 160,00 rpm at 4°C for 30 min.

The resulting pellet was resuspended in 50 ml STET buffer (50 mM Tris-HCl, pH 8.5, 8% saccharose, 5% triton X-100, 50 mM EDTA) and the suspension was sonicated (3 x 45 sec) and centrifuged at 30,000 rpm for 20 min. The described washing procedure was repeated twice and twice again with 50 ml of 50 mM Tris-HCl, pH 8.5, and 100 mM NaCl.

Denaturation

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The sample was resuspended in 30 ml buffer A (50 mM Tris-HCl, pH 8.0, 6 M Gu-HCl, 5 mM imidazole) and incubated at room temperature for at least 2 h.

Immunotoxin purification by IMAC

The sample was centrifuged at 120,00 rpm for 30 min. The supernatant was analysed by chelated metal affinity chromatography (IMAC) using 20 ml Ni⁺⁺-filled chelating sepharose FF resin (Pharmacia).

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The column was washed with 5 vol water, loaded with 5 vol of 0.1 M NiSO₄, washed with 5 vol water and equalized with 5 vol buffer A.

Once the sample had been injected, the column was washed with 5 vol buffer A. Adsorbed proteins were eluted in step of pH using the following buffers:

5 50 mM Tris-acetate, pH 5.5; 6 M Gu-HCl (Buffer B);

50 mM Tris-acetate, pH 4.0; 6 M Gu-HCl (Buffer C).

The immunotoxin was eluted in buffer C.

Immunotoxin reduction

The sample obtained from IMAC (in a concentration of 1-2 mg/ml) was brought to pH 8.3 with 1 M Tris base. 2 mM EDTA and final 300 mM DTT were added. The resulting product was incubated at room temperature for 3 h.

Immunotoxin refolding

The reduced sample was rapidly diluted (1:100) in the refolding buffer (50 mM Tris-HCI, pH 8.3, 0.5 M L-Arg, 2 mM EDTA, 4 mM GSSG, 2 mM DTT) and incubated at 10°C for 60 h.

Immunotoxin concentration and dialysis

The sample (ca. 500 ml) was added with Tween-20 (final 0.005%) and concentrated by ultrafiltration through membrane Amicon YM 10.

Dialysis was carried out using a 10,000-cut-off membrane vs dialysis buffer (50 mM Tris-HCl, pH 8.0, 0.2 M NaCl, 0,005% Tween-20, 10% glycerol).

The resulting product was centrifuged at 120,00 rpm for 30 min and the supernatant containing the immunotoxin was recovered.

Inhibition of cell protein synthesis (chemical immunoconjugate)

The capacity of clavin and of immunotoxin Mgr6-clavin for inhibiting the protein synthesis was measured on SKBr3 (ErbB2⁺ cells) and on MeWo (ErbB2⁻ cells) cultured in RPMI 1640 containing 10% FCS. The test was carried out essentially as described by Casalini *et al.*, 1993.

The immunotoxin, the toxin or the monoclonal antobody were diluted in turn in the culture medium. Cells (1.2 x 10⁶) were incubated at 4°C for 3 h in polypropylene test tubes, in 800 ul culture medium containing the appropriate concentrations of immunotoxin, toxin or monoclonal antibody alone. Control cells were incubated

only with the culture medium. Cells were then centrifuged, resuspended in a fresh culture medium and seeded in triplicate in 96-well plates (3×10^5 cells/well).

After incubation at 37°C for 48 h, the culture medium was removed and a fresh culture medium containing [³H] proline (1uCi/well) was added. 48 h later, cells were washed and the amount of [³H] proline incorporated was determined.

In various tests, clavin shows a dose/response effect with IC $\underline{50}$ values ranging from 0.1 to 1 μ M.

The cytotoxicity of the Mgr6-clavin conjugate is similar to that of ricin A bound to the same monoclonal antibody.

Inhibition of cell protein synthesis (recombinant immunoconjugate)

The capacity of recombinant immunotoxin Mgr6-clavin for inhibiting the protein synthesis was measured as already described for the chemical immunoconjugate. The IC $\underline{50}$ of recombinant immunotoxin ranges from 0.1 to 1 μ M, whereas antibody Mgr6 does not produce any effect. Therefore, clavin is a promising candidate for immunotoxin production.

SEQUENCE LISTING

(1) GENERAL INFORMATION:

- (i) APPLICANT:
 - (A) NAME: MINISTERO UNIVERSITA' RICERCA SCIENTIFICA E TECNOLOGICA
 - (B) STREET: c/o NOTARBARTOLO & GERVASI S.P.A. Corso di Porta Vittoria, 9
 - (C) CITY: MILAN
 - (D) STATE: MI
 - (E) COUNTRY: ITALY
 - (F) POSTAL CODE (ZIP): 20122
 - (G) TELEPHONE: 02-5417991
 - (H) TELEFAX: 02-54179920
- (ii) TITLE OF INVENTION: A protein capable of inhibiting ribosomal activity, its preparation and use as a chemical or recombinant immunoconjugate, and the cDNA sequence expressing said protein.
- (iii) NUMBER OF SEQUENCES: 16
- (iv) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Floppy disk
 - (B) COMPUTER: IBM PC compatible
 - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 - (D) SOFTWARE: PatentIn Release #1.0, Version #1.30 (EPO)
- (vi) PRIOR APPLICATION DATA:
 - (A) APPLICATION NUMBER: IT FI96A000155
 - (B) FILING DATE: 27-JUN-1996
- (2) INFORMATION FOR SEO ID NO: 1:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1011 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: cDNA
 - (iii) HYPOTHETICAL: NO
 - (iv) ANTI-SENSE: NO
 - (vi) ORIGINAL SOURCE:
 - (A) ORGANISM: Aspergillus clavatus
 - (B) STRAIN: IFO8605

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 1:

60	GGAAAAGAAA	CAGAACGCTT	TGTGAGAAAC	TGGTGAGATT	AAAGGATATG	AACCAGAAAC
120	AGATCCAACG	TCTGACTCAG	GAGGATATTG	CACCATCGAT	GAAAAGTAAT	ACĄAAAGAGA
180	CAATGGAGTC	CGAGCTGGGT	TCGCCCACAT	ATGCTTCAAG	CAACTTCGGA	AAATAATAGT
240	CCCAGGAAAA	TCGCGGTTCT	TGCTAGATCC	ATATAAAAGC	GCCAGAGCAC	TCTCGAGTCA
300	CAAGAACCTC	TGGTCGCAAT	ACATCCAAAA	CATCTTAACC	TGATCTCAAG	CCCAAGATCG
360	CGAGGAGCGC	CTTCGCCTCT	CTTGCGATGC	CGTGACCGCC	CCCTCACGGC	GTCCTGGTGG
420	GTATGAGAAC	AGACCAACAA	AAGAACCCAA	GAACGAGCAG	GGACTTGCAT	GCGGCGACCT
480	GCCTCTCTCC	CCCACCACGC	GAGAGCAACG	GAACAATGCC	TCTACAACCA	AAGCGCCTCC
540	CGGCGATGGA	ACGGCTACGA	TGGTTCACCA	CTATCCTCAC	CCGGTAGCAG	GACGGCAAGA
600	CCGCCCTCCC	CGGACTGCGA	TGGGGAAATT	GCCCATCAAG	AGGGCCGCAC	AAGATCCTCA
660	CCCAACATTC	TGCTGGAGTT	GACCATTACC	TGGCAAGAAT	AGAATGGTGA	AAGCACAGCA
720	CGGCCCGGCA	AGGAGGACCC	AAGAAGCCCA	TTTCGACTCG	ACCAGTATAA	CCCGATGGAC
780	CACGAGGGAG	TTGTTGCCCA	TTCTGCGGCA	TAACAAGGTG	ACACCTATCC	CGGGTCATCT
840	TATAGTTTGC	TGCACAGGGA	TAAATGGGCT	CTGCTCTCAT	ACCTGAAGCT	AACCAGGGTG
900	GGGAGGGAGC	TGGGAAGTGG	TATATCGCAT	CGGCTGATAC	TCTTCAACCA	CATTGGTCGT
960	ATCCCAGTCT	TATCTAGTCA	TGTTCTATGT	GTGCAGAACT	ACATATGTTG	TGAATGTTTC
1011	A	ааааааааа	CATTGCAAAA	TATTGCACTT	TATCTATGCA	CTCGCTTTGA

(2) INFORMATION FOR SEQ ID NO: 2:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 453 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (vi) CRIGINAL SOURCE:
 - (A) ORGANISM: Aspergillus clavatus (B) STRAIN: IF08605

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 2:

GCGGCGACCT	GGACTTGCAT	GAACGAGCAG	AAGAACCCAA	AGACCAACAA	GTATGAGAAC	60
AAGCGCCTCC	TCTACAACCA	GAACAATGCC	GAGAGCAACG	CCCACCACGC	GCCTCTCTCC	120
GACGGCAAGA	CCGGTAGCAG	CTATCCTCAC	TGGTTCACCA	ACGGCTACGA	CGGCGATGGA	180
AAGATCCTCA	AGGGCCGCAC	GCCCATCAAG	TGGGGAAATT	CGGACTGCGA	CCGCCCTCCC	240
AAGCACAGCA	AGAATGGTGA	TGGCAAGAAT	GACCATTACC	TGCTGGAGTT	CCCAACATTC	300
CCĊGATGGAC	ACCAGTATAA	TTTCGACTCG	AAGAAGCCCA	AGGAGGACCC	CGGCCCGGCA	360
CGGGTCATCT	ACACCTATCC	TAACAAGGTG	TTCTGCGGCA	TTGTTGCCCA	CACGAGGGAG	420
AACCAGGGTG	ACCTGAAGCT	CTGCTCTCAT	TAA			453

(2) INFORMATION FOR SEQ ID NO: 3:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 150 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS:
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (iii) HYPOTHETICAL: NO
- (vi) ORIGINAL SOURCE:
 - (A) ORGANISM: Aspergillus clavatus
 - (B) STRAIN: IF08605
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 3:
- Ala Ala Thr Trp Thr Cys Met Asn Glu Gln Lys Asn Pro Lys Thr Asn 1 5 10
- Lys Tyr Glu Asn Lys Arg Leu Leu Tyr Asn Gln Asn Asn Ala Glu Ser 20 25 30
- Asn Ala His His Ala Pro Leu Ser Asp Gly Lys Thr Gly Ser Ser Tyr 35 40
- Pro His Trp Phe Thr Asn Gly Tyr Asp Gly Asp Gly Lys Ile Leu Lys 50 55
- Gly Arg Thr Pro Ile Lys Trp Gly Asn Ser Asp Cys Asp Arg Pro Pro 65 70 75 80
- Lys His Ser Lys Asn Gly Asp Gly Lys Asn Asp His Tyr Leu Leu Glu 85 90 95
- Phe Pro Thr Phe Pro Asp Gly His Gln Tyr Asn Phe Asp Ser Lys Lys 100 105 110
- Pro Lys Glu Asp Pro Gly Pro Ala Arg Val Ile Tyr Thr Tyr Pro Asn 115 120 125
- Lys Val Phe Cys Gly Ile Val Ala His Thr Arg Glu Asn Gln Gly Asp 130 135 140
- Leu Lys Leu Cys Ser His
 - 45 15

WO 97/49726 13

(2) INFORMATION FOR SEQ ID NO: 4:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1314 base pairs

PCT/EP97/03359

- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (vi) ORIGINAL SOURCE:
 - (A) ORGANISM: Aspergillus clavatus
 - (B) STRAIN: IFO8605

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 4:

ATGCGGGGTT	CTCATCATCA	TCATCATCAT	GGTATGGCTA	GCATGACTGG	TGGACAGCAA	60
ATGGGTCGGG	ATCTGTACGA	CGATGACGAT	AAGGATCGAT	GGGGATCCCA	GGTGCAGCTG	120
CAGGAGTCTG	GGGCAGAGCT	TGTGAAGCCA	GGGGCCTCAG	TCAAGTTGTC	CTGCACAGCT	180
TCTGGCTTCA	ACATTAAAGA	CACCTATATG	CACTGGGTGA	AGCAGAGGCC	TGAACAGGGC	240
CTGGAGTGGA	TTGGAAGGAT	TGATCCTGCG	AATGGTAATA	CTAAATATGA	CCCGAAGTTC	300
CAGGGCAAGG	CCACTATAAC	AGCAGACACA	TCCTCCAACA	CAGCCTACCT	GCAGCTCAGC	360
AGCCTGACAT	CTGAGGACAC	TGCCGTCTAT	TACTGTGCTA	GAGGAGAATA	TGATTATCCT	420
TTTCCTTACT	GGGGCCAAGG	GACCTCGGTC	ACCGTCTCCT	CAGGTGGAGG	CGGTTCAGGC	480
GGAGGTGGCT	CTGGCGGTGG	CGGATCGTAC	ATCGAGCTCA	CTCAGTCTCC	AGCTTCCTTA	540
GCTGTATCTC	TGGGGCAGAG	GGCCACCATC	TCATGCAGGG	CCAGCCAAAG	TGTCAGTACA	600
TCTAGGTATA	GTTATATGCA	CTGGTACCAA	CAGAAACCAG	GACAGCCACC	CAAACTCCTC	660
ATCAAGTATG	CATCCAACCT	AGAATCTGGG	GTCCCTGCCA	GGTTCAGTGG	CAGTGGGTCT	720
GGGACAGACT	TCACCCTCAA	CATCCATCCT	GTGGAGGAGG	AGGATACTGC	AACATATTAC	780
TGTCAGCACA	GTTGGGAGAT	TCCTCGGACG	TTCGGTGGAG	GGACCAAGCT	GGAGCTGAAA	840
CGGGCGGGAT	CCCCGGAATT	CGCAGCGACC	TGGACTTGCA	TGAACGAGCA	GAAGAACCCA	900
AAGACCAACA	AGTATGAGAA	CAAGCGCCTC	CTCTACAACC	AGAACAATGC	CGAGAGCAAC	960
GCCCACCACG	CGCCTCTCTC	CGACGGCAAG	ACCGGTAGCA	GCTATCCTCA	CTGGTTCACC	1020
AACGGCTACG	ACGGCGATGG	AAAGATCCTC	AAGGGCCGCA	CGCCCATCAA	GTGGGGAAAT	1080
TCGGACTGCG	ACCGCCCTCC	CAAGCACAGC	AAGAATGGTG	ATGGCAAGAA	TGACCATTAC	1140
CTGCTGGAGT	TCCCAACATT	CCCCGATGGA	CACCAGTATA	ATTTCGACTC	GAAGAAGCCC	1200
AAGGAGGACC	CCGGCCCGGC	ACGGGTCATC	TACACCTATC	CTAACAAGGT	GTTCTGCGGC	1260
ATTGTTGCCC	ACACGAGGGA	GAACCAGGGT	GACCTGAAGC	TCTGCTCTCA	TTAG	1314

- (2) INFORMATION FOR SEQ ID NO: 5:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 437 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS:
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (iii) HYPOTHETICAL: NO
 - (vi) ORIGINAL SOURCE:
 - (A) ORGANISM: Aspergillus clavatus
 - (B) STRAIN: IFO8605
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 5:
 - Met Arg Gly Ser His His His His His Gly Met Ala Ser Met Thr

 1 5 10 15
 - Gly Gly Gln Met Gly Arg Asp Leu Tyr Asp Asp Asp Asp Lys Asp 20 25 30
 - Arg Trp Gly Ser Gln Val Gln Leu Gln Glu Ser Gly Ala Glu Leu Val 35 40 45
 - Lys Pro Gly Ala Ser Val Lys Leu Ser Cys Thr Ala Ser Gly Phe Asn 50 55 60
 - Ile Lys Asp Thr Tyr Met His Trp Val Lys Gln Arg Pro Glu Gln Gly 65 70 75 80
 - Leu Glu Trp Ile Gly Arg Ile Asp Pro Ala Asn Gly Asn Thr Lys Tyr 85 90 95
 - Asp Pro Lys Phe Gln Gly Lys Ala Thr Ile Thr Ala Asp Thr Ser Ser 100 105 110
 - Asn Thr Ala Tyr Leu Gln Leu Ser Ser Leu Thr Ser Glu Asp Thr Ala 115 120 125
 - Val Tyr Tyr Cys Ala Arg Gly Glu Tyr Asp Tyr Pro Phe Pro Tyr Trp 130 135 140
 - Gly Gln Gly Thr Ser Val Thr Val Ser Ser Gly Gly Gly Gly Ser Gly 145 150 155 160
 - Gly Gly Gly Ser Gly Gly Gly Ser Tyr Ile Glu Leu Thr Gln Ser 165 170 175
 - Pro Ala Ser Leu Ala Val Ser Leu Gly Gln Arg Ala Thr Ile Ser Cys 180 185 190
 - Arg Ala Ser Gln Ser Val Ser Thr Ser Arg Tyr Ser Tyr Met His Trp
 195 200 205
 - Tyr Gln Gln Lys Pro Gly Gln Pro Pro Lys Leu Leu Ile Lys Tyr Ala 210 215 220
 - Ser Asn Leu Glu Ser Gly Val Pro Ala Arg Phe Ser Gly Ser Gly Ser 225 230 235 240
 - Gly Thr Asp Phe Thr Leu Asn Ile His Pro Val Glu Glu Glu Asp Thr 245 250 255

Ala Thr Tyr Tyr Cys Gln His Ser Trp Glu Ile Pro Arg Thr Phe Gly 260 265 270

Gly Gly Thr Lys Leu Glu Leu Lys Arg Ala Gly Ser Pro Glu Phe Ala 275 280 285

Ala Thr Trp Thr Cys Met Asn Glu Gln Lys Asn Pro Lys Thr Asn Lys 290 295 300

Tyr Glu Asn Lys Arg Leu Leu Tyr Asn Gln Asn Asn Ala Glu Ser Asn 305 310 315 320

Ala His His Ala Pro Leu Ser Asp Gly Lys Thr Gly Ser Ser Tyr Pro 325 330 335

His Trp Phe Thr Asn Gly Tyr Asp Gly Asp Gly Lys Ile Leu Lys Gly 340 345 350

Arg Thr Pro Ile Lys Trp Gly Asn Ser Asp Cys Asp Arg Pro Pro Lys 355 360 365

His Ser Lys Asn Gly Asp Gly Lys Asn Asp His Tyr Leu Leu Glu Phe 370 380

Pro Thr Phe Pro Asp Gly His Gln Tyr Asn Phe Asp Ser Lys Lys Pro 395 390 395

Lys Glu Asp Pro Gly Pro Ala Arg Val Ile Tyr Thr Tyr Pro Asn Lys 405 410 415

Val Phe Cys Gly Ile Val Ala His Thr Arg Glu Asn Gln Gly Asp Leu 420 425 430

Lys Leu Cys Ser His 435

(2) INFORMATION FOR SEQ ID NO: 6:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 28 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 6:

ACGTAAGCTT CTAATGAGAG CAGAGCTT

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

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16
(2) INFORMATION FOR SEQ ID NO: 7:
     (i) SEQUENCE CHARACTERISTICS:
          (A) LENGTH: 31 base pairs
          (B) TYPE: nucleic acid
          (C) STRANDEDNESS: single
          (D) TOPOLOGY: linear
    (ii) MOLECULE TYPE: DNA (genomic)
   (iii) HYPOTHETICAL: NO
   (iv) ANTI-SENSE: NO
    (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 7:
ACGTCTGCAG TGACCTGGAC CTGCTTGAAC G
(2) INFORMATION FOR SEQ ID NO: 8:
     (i) SEQUENCE CHARACTERISTICS:
         (A) LENGTH: 35 base pairs
         (B) TYPE: nucleic acid
          (C) STRANDEDNESS: single
         (D) TOPOLOGY: linear
    (ii) MOLECULE TYPE: DNA (genomic)
   (iii) HYPOTHETICAL: NO
   (iv) ANTI-SENSE: NO
    (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 8:
(2) INFORMATION FOR SEQ ID NO: 9:
    (i) SEQUENCE CHARACTERISTICS:
         (A) LENGTH: 17 base pairs
          (B) TYPE: nucleic acid
         (C) STRANDEDNESS: single
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(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 9:	
GACTCGAGTC GACATCG	17
(2) INFORMATION FOR SEQ ID NO: 10:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 24 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
(ii) MOLECULE TYPE: DNA (genomic)	
(iii) HYPOTHETICAL: NO	
(iv) ANTI-SENSE: NO	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 10:	
ACGTGGATCC TCTACAACCA GAAC	24
(2) INFORMATION FOR SEQ ID NO: 11:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 17 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
(ii) MOLECULE TYPE: DNA (genomic)	
(iii) HYPOTHETICAL: NO	
(iv) ANTI-SENSE: NO	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 11:	
TGAACCAGTG AGGATAG	17
(2) INFORMATION FOR SEQ ID NO: 12:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 25 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: DNA (genomic)	
(iii) HYPOTHETICAL: NO	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 12:	
ACGTCTGCAG GCGCTTGTTC TCATA	25

(2) INFO	RMATION FOR SEQ ID NO: 13:	
(i)	SEQUENCE CHARACTERISTICS: (A) LENGTH: 51 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(ii)	MOLECULE TYPE: DNA (genomic)	
(iii)	HYPOTHETICAL: NO	
(xi)	SEQUENCE DESCRIPTION: SEQ ID NO: 13:	
GATCCTGCA	AG CGACCTGGAC TTGCATGAAC GAGCAGAAGA ACCCAAAGAC C	51
(2) INFOR	RMATION FOR SEQ ID NO: 14:	
(i)	SEQUENCE CHARACTERISTICS: (A) LENGTH: 28 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(ii)	MOLECULE TYPE: DNA (genomic)	
(iii)	HYPOTHETICAL: NO	
(iv)	ANTI-SENSE: NO	
(xi)	SEQUENCE DESCRIPTION: SEQ ID NO: 14:	
ACGTAAGCT	TT CTAATGAGAG CAGAGCTT	28
(2) INFOR	RMATION FOR SEQ ID NO: 15:	
(i)	SEQUENCE CHARACTERISTICS: (A) LENGTH: 24 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(ii)	MOLECULE TYPE: DNA (genomic)	
(iii)	HYPOTHETICAL: NO	
(iv)	ANTI-SENSE: NO	
(xi)	SEQUENCE DESCRIPTION: SEQ ID NO: 15:	
AATTCGATC	CG AAGGTCGTAC TGCA	24
(2) INFOR	RMATION FOR SEQ ID NO: 16:	
(i)	SEQUENCE CHARACTERISTICS: (A) LENGTH: 16 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	

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(ii) MOLECULE TYPE: DNA (genomic)

- (iii) HYPOTHETICAL: NO
 - (iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 16:

GTACGACCTT CGATCG

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Claims

- Nucleotide sequence SEQ ID No: 2.
- 2. Protein capable of inactivating the ribosomal activity, the amino acid sequence
- 2 SEQ ID No: 3.
- 1 3. Nucleotide sequence SEQ ID No: 4, encoding for the Mgr6-clavin
- 2 immunotoxin.
- 1 4. Protein sequence SEQ ID No: 5 of Mgr6-clavin immunotoxin.
- 1 5. Conjugates, obtained by chemical conjugation or by genic recombination, of the
- 2 protein as claimed in claim 2 with hormones, liposomes, monoclonal antibodies,
- 3 growth factors, cytokines, transferrin and peptides consisting of fragments of said
- 4 proteins.
- 1 6. The conjugates as claimed in claim 5, wherein the protein is conjugated with
- 2 monoclonal antibodies.
- 7. The conjugates as claimed in claim 5, wherein the monoclonal antibody is
- 2 Mgr6.
- 8. Pharmaceutical compositions containing, as active ingredient, the protein as
- claimed in claim 2 and/or the conjugates as claimed in any of claims 5 to 7,
- 3 combined with suitable additives.
- 9. Use of the protein as claimed in claim 2 and/or of the conjugates as claimed in
- 2 any of claims 5 to 7, or mixtures thereof, for the preparation of pharmaceutical
- 3 preparations useful as anticancer and/or antiviral agents. A protein capable of
- 4 inhibiting ribosomal activity, its preparation and use as a chemical or recombinant
- 5 immunoconjugate, and the cDNA sequence expressing said protein.

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A. CLASSIFICATION OF SUBJECT MATTER 1PC 6 C07K14/38 C07H C07K19/00 A61K47/48 C12N15/62 According to International Patent Classification (IPC) or to both national classification and IPC **B. FIELDS SEARCHED** Minimum documentation searched (classification system followed by classification symbols) IPC 6 CO7K C12N A61K Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched Electronic data base consulted during the international search (name of data base and, where practical, search terms used) C. DOCUMENTS CONSIDERED TO BE RELEVANT Citation of document, with indication, where appropriate, of the relevant passages Relevant to claim No Х EMBL/GENEBANK/DDBJ databases 1,2 Accesion number U19383, 4 July 1995 Parente et al. "Clavin, type 1 ribosome-inactivating protein from Aspergillus clavatus IF8605" XP002047873 see abstract 3-9 P.X & PARENTE ET AL., : "Clavin, a type 1 1.2 ribosome-inactivating protein from Aspergillus clavatus IFO 8605" EUROPEAN JOURNAL OF BIOCHEMISTRY, vol. 239, - 15 July 1996 BERLIN, DE, pages 272-280, See the whole document and specially figure 1 -/--Further documents are listed in the continuation of box C Patent family members are listed in annex Special categories of cited documents "T" later document published after the international filing date or priority date and not in conflict with the application but *A* document defining the general state of the art which is not considered to be of particular relevance cited to understand the principle or theory underlying the *E" earlier document but published on or after the international "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to filing date "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another involve an inventive step when the document is taken alone "Y" document of particular relevance; the claimed invention citation or other special reason (as specified) cannot be considered to involve an inventive, step when the "O" document referring to an oral disclosure, use, exhibition or document is combined with one or more other suc ments, such combination being obvious to a person skilled document published prior to the international filing date but later than the priority date claimed "&" document member of the same patent family Date of the actual completion of the international search Date of mailing of the international search report 0 9 -12- 1997 24 November 1997 Name and mailing address of the ISA Authorized officer European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijewijk Tel. (+31-70) 340-2040, Tx. 31 651 epo nl, Mateo Rosell, A.M. Fax: (+31-70) 340-3016

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